

Low Phytate Soybeans: *Can we solve the germ problem?*

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Low Phytate Soybean Germplasm

Mutations	Germplasm	Phytate content*	Germplasm Citation
<i>Lpa1</i> & <i>Lpa2</i>	Cx1834	37% normal	Wilcox et al. (2000)
<i>MIPS1</i>	V99-5089 Lr33 <i>GM-Lpa-TW1</i>	67% normal	Saghai Maroof & Buss (2008) Hitz et al. (2002) Yuan et al. (2007)

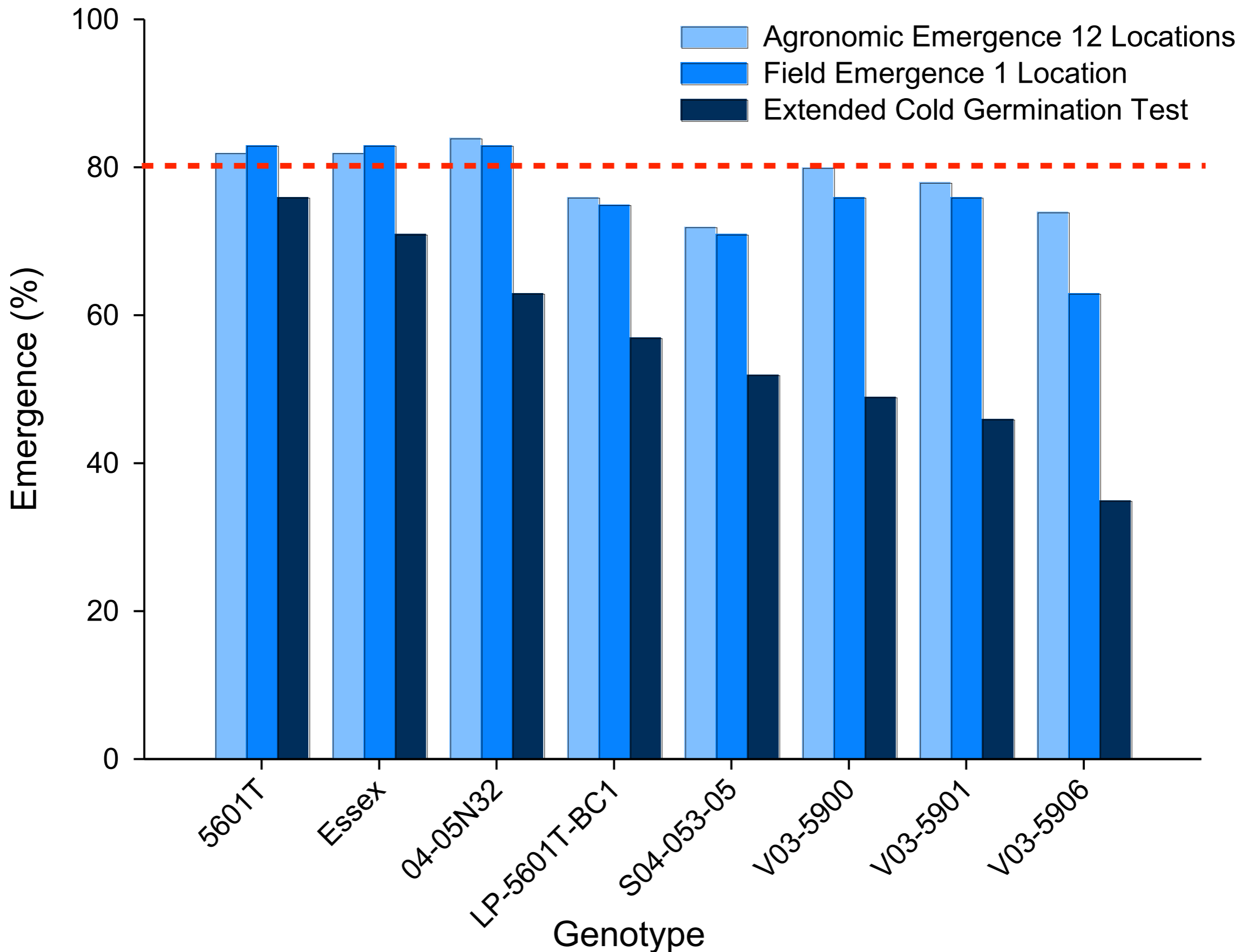
* Summarized from Maupin (2010)

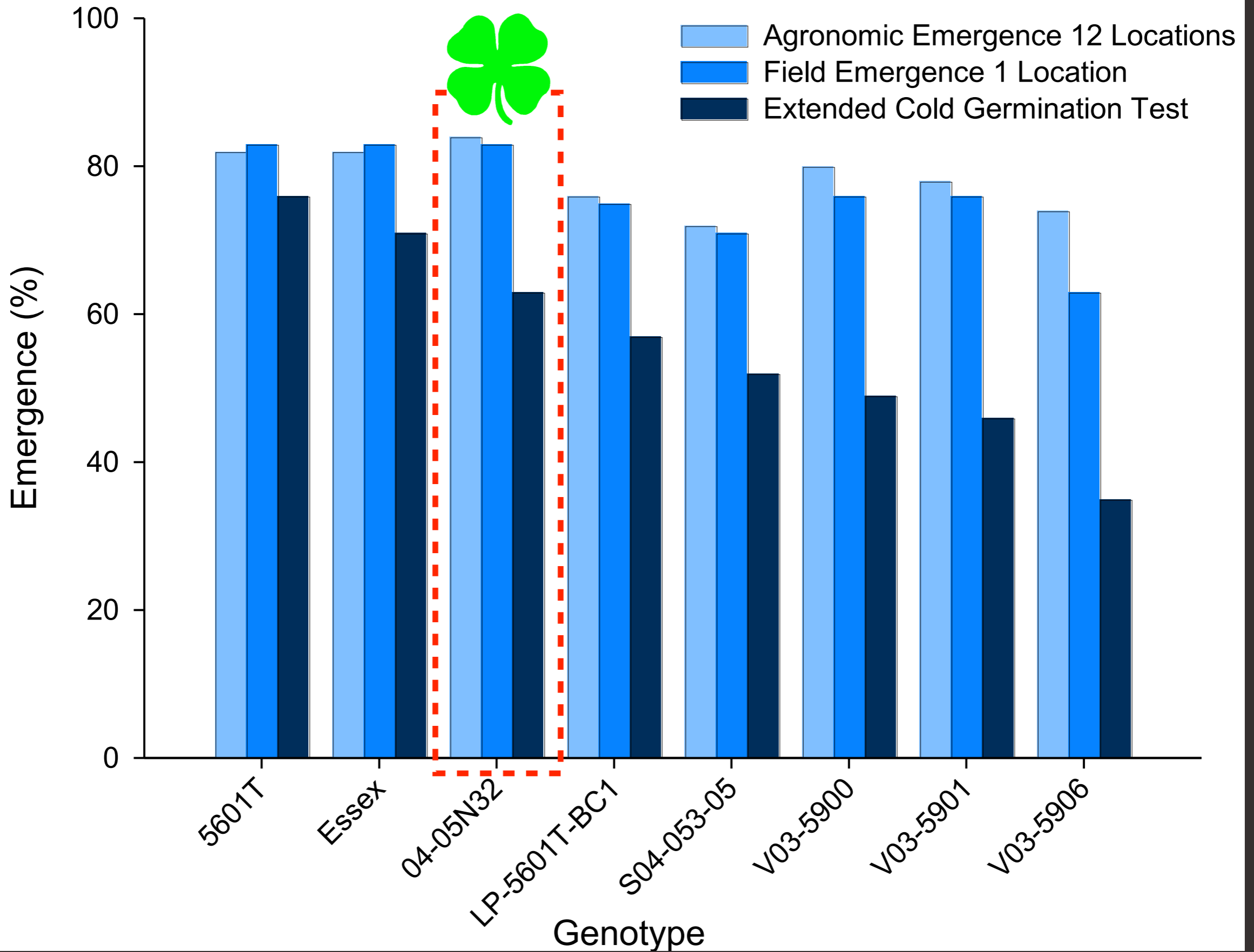
I2 Environment Study: Experiment

- Germplasm
 - 3 *Lpa* mutants
 - 3 *MIPSI* mutants
 - 2 WT
- 12 Environments
 - 2 years x 6 locations
Queenstown, MD, Portageville, MO, Plymouth, NC,
Knoxville, TN, Blacksburg, VA, and Mount Holly, VA in
2008 and 2009.
- Data
 - P content: Pi, HPIC, and new colorimetric phytate
 - Germination & Emergence
 - stand counts
 - extended cold germination test (ECGT)
 - warm germination *Trimble & Fehr, 2010*
 - Field emergence: seed from 12 env planted into 1 env



Maupin 2010





I2 Environment Study: Emergence Results

	Standard Germination	2009 Agronomic Emergence						Standard Germination	2010 Agronomic Emergence						
Genotype	VAMH 2008 Seed	NC	MD	VAMH	TN	VABB	MO	VAMH 2009 Seed	MD	NC	VABB	TN	MO	VAMH	Mean [§]
5601T	92	98	96	83	88	90	47	98	95	87	85	75	79	56	82ab
Essex	84	95	95	88	82	89	53	97	93	86	82	74	76	69	82ab
04-05N32	94	97	93	89	91	88	69	98	96	85	76	75	78	67	84a
LP-5601T-BC1	83	95	91	91	85	70	50	95	88	82	69	68	56	66	76cde
S04-053-05	88	95	71	86	80	58	42	96	94	82	72	77	65	46	72e
V03-5900	83	96	96	89	87	86	39	97	97	83	80	72	75	56	80abc
V03-5901	87	97	93	79	84	87	32	98	97	85	75	75	71	68	78bcd
V03-5906	84	95	91	80	83	80	30	97	95	76	71	66	72	51	74de
Mean [§]		96a	91abc	85bcd	85bcd	81cde	45g		94ab	83cd	76de	73e	71e	60f	
Correlation															
Standard Germination															
<i>r</i>		0.74	-0.05	-0.11	0.56	0.22	0.54		0.82	0.43	0.61	0.42	0.89	0.20	
<i>P</i>		*	ns	ns	ns	ns	ns		**	ns	ns	ns	**	ns	
Extended Cold Germination Test															
<i>r</i>		0.34	0.16	0.56	0.36	0.20	0.80		0.21	0.63	0.66	0.77	0.33	-0.18	
<i>P</i>		ns	ns	ns	ns	ns	*		ns	ns	ns	*	ns	ns	
Field Emergence Test															
<i>r</i>		0.53	0.45	0.40	0.49	0.57	0.57		0.09	0.74	0.69	0.40	0.27	0.39	
<i>P</i>		ns	ns	ns	ns	ns	ns		ns	*	ns	ns	ns	ns	

I2 Environment Study: Other Notable Results

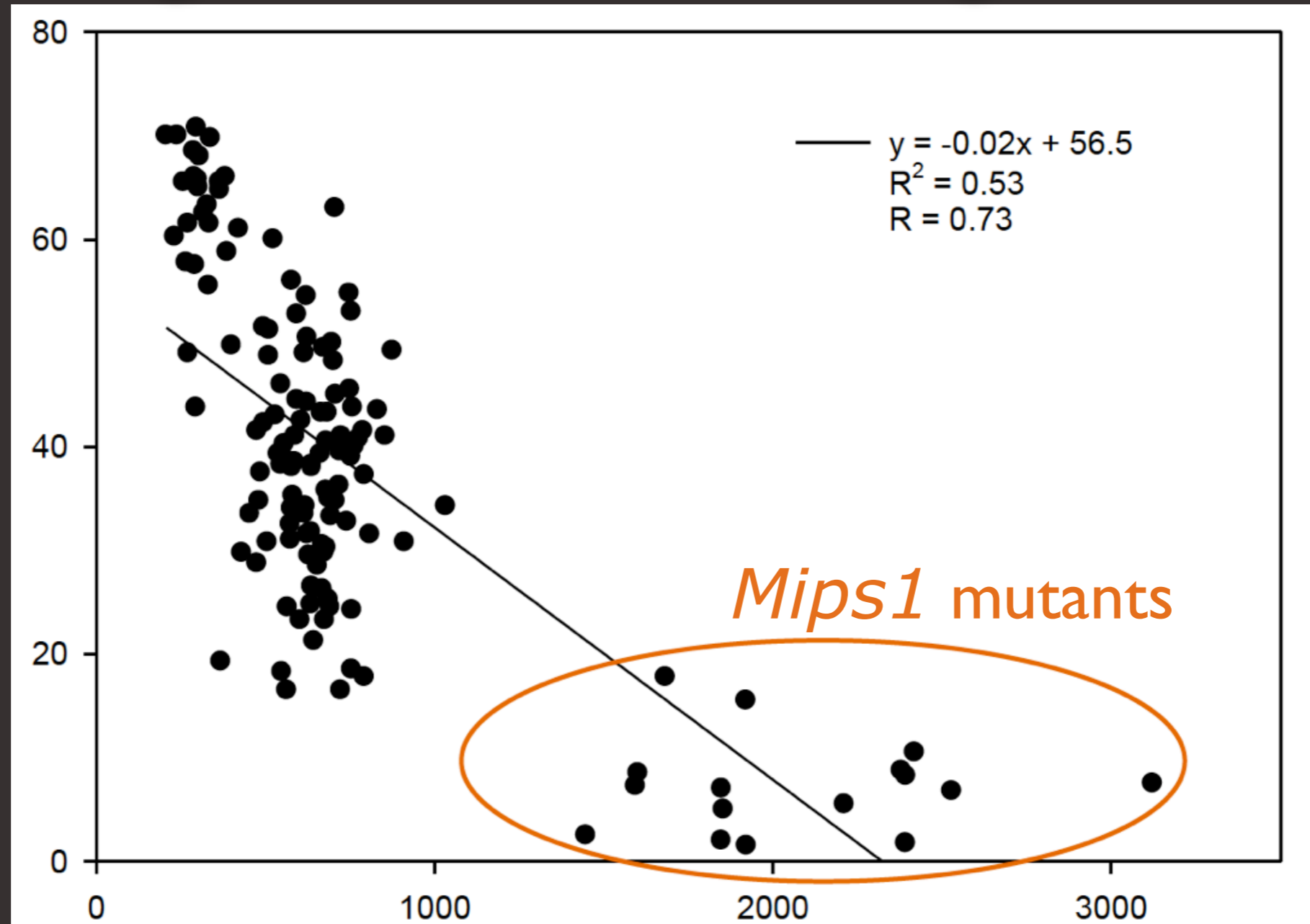
- ▶ Low phytate trait *per se* not associated with poor agronomic performance
- ▶ Significant differences for P content among *Lpa* mutant lines
- ▶ Differential stability for P content
- ▶ Low overall P = good emergence

Low phytate soybeans can have poor germination and emergence.



Low phytate soybeans can have poor germination and emergence.

F_{2.4} % Emergence



F₂ Inorganic Phosphorus (ug g⁻¹)

Low phytate soybeans can have poor germination and emergence.

CX1834-derived:

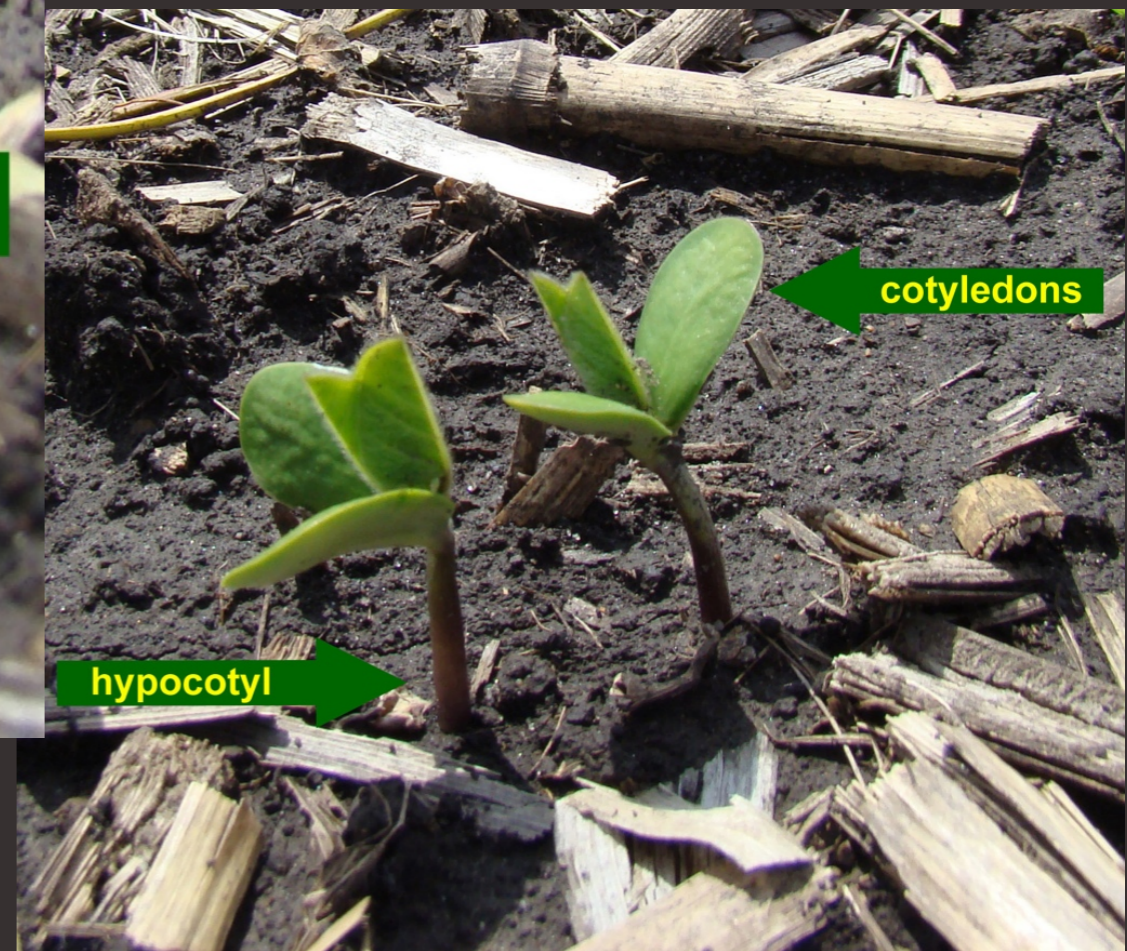
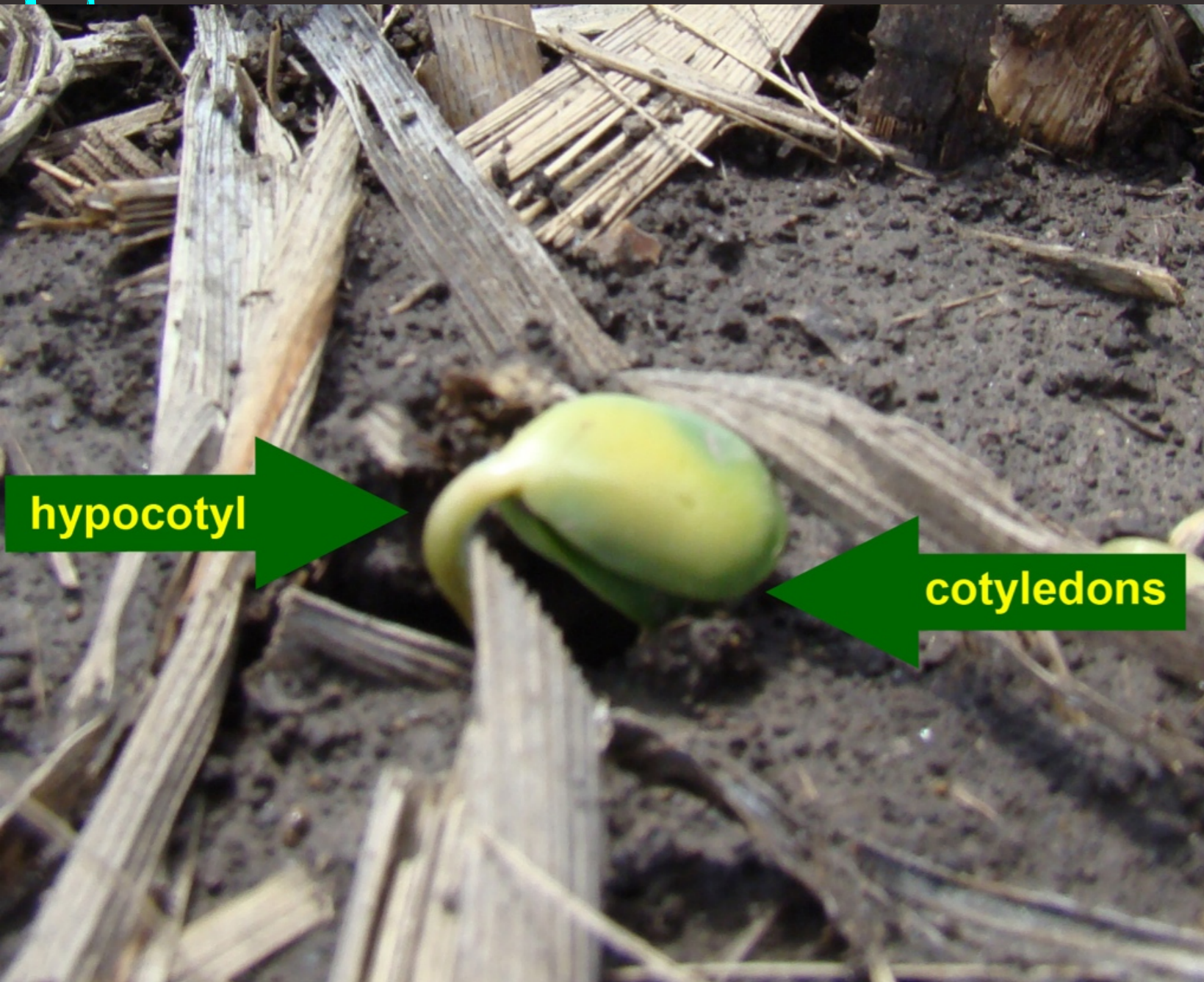
Hulke et al. (2004)

Oltmans et al. (2005)

Spear and Fehr (2007)

Scaboo et al. (2009)

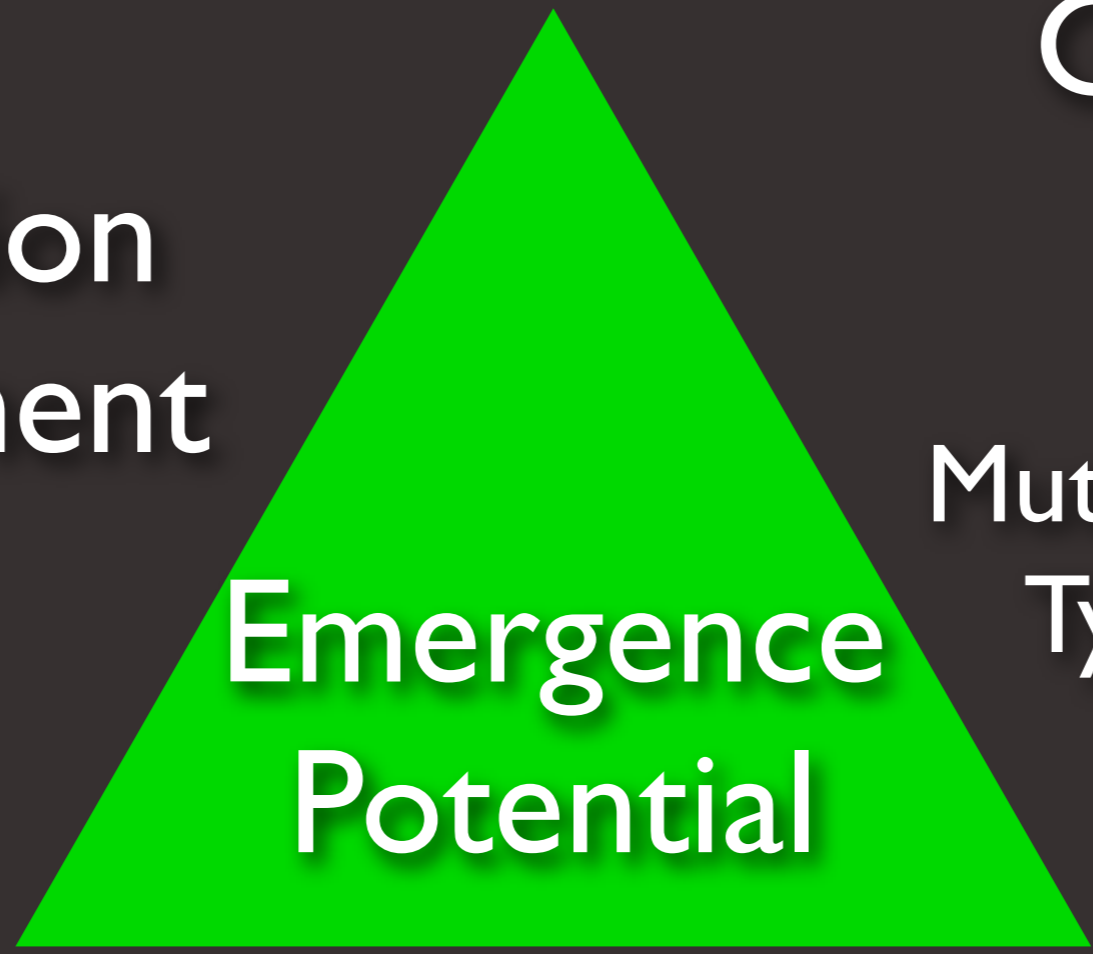
Germination is Not Field Emergence



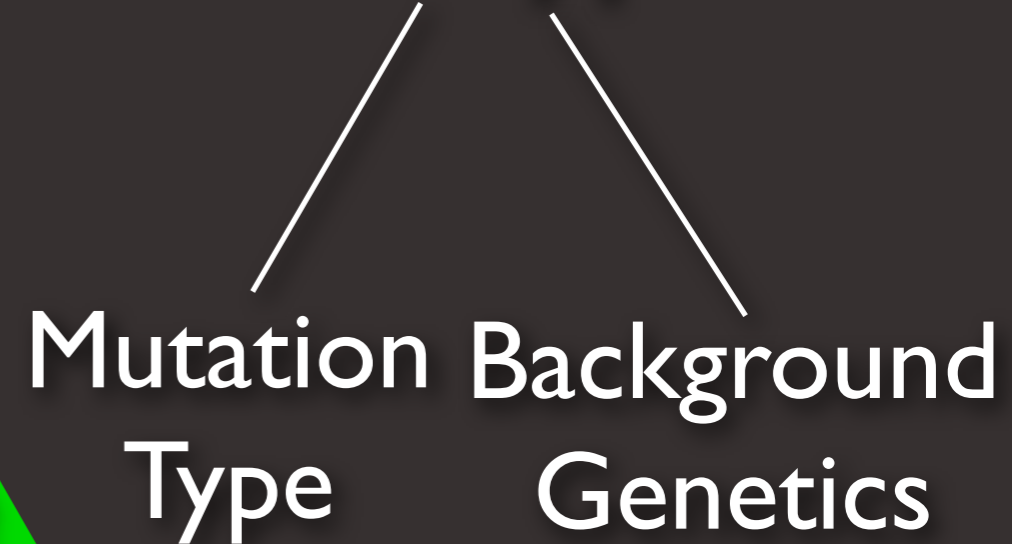
<http://extension.entm.purdue.edu/pestcrop/2010/issue7/index.html>

Emergence is a Dynamic Trait

Seed
Production
Environment

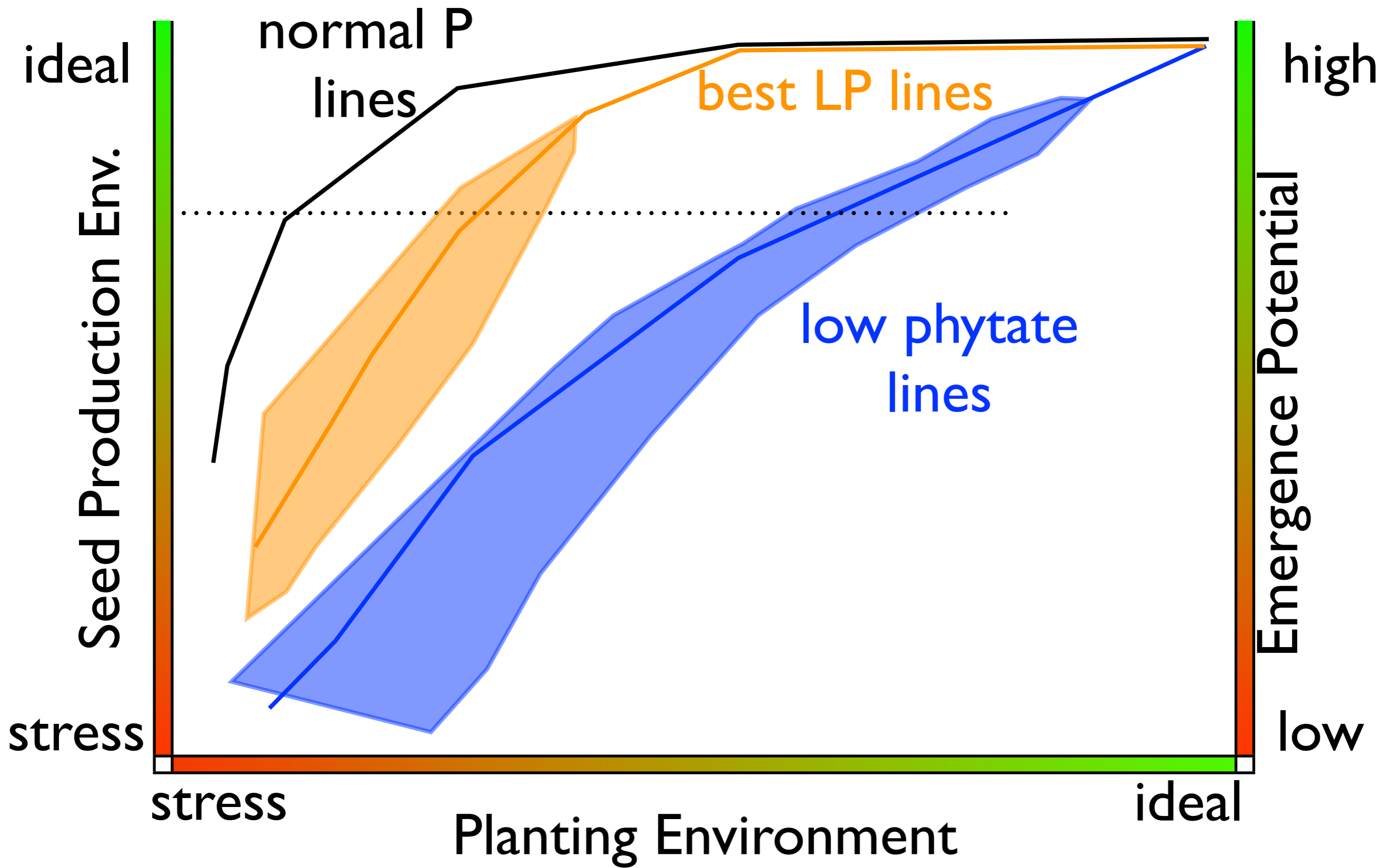


Genotype



Planting
Environment

Emergence Model



Theor Appl Genet (2009) 118:1211–1221

DOI 10.1007/s00122-009-0975-8

ORIGINAL PAPER

Isolation and characterisation of an *lpa* (low phytic acid) mutant in common bean (*Phaseolus vulgaris* L.)

**Bruno Champion · Francesca Sparvoli · Enrico Doria ·
Giovanni Tagliabue · Incoronata Galasso ·
Marzia Fileppi · Roberto Bollini · Erik Nielsen**

*Common assumptions about germination
and emergence in low phytate soybeans.*

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natural selection

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LP populations = typical pops

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linkage

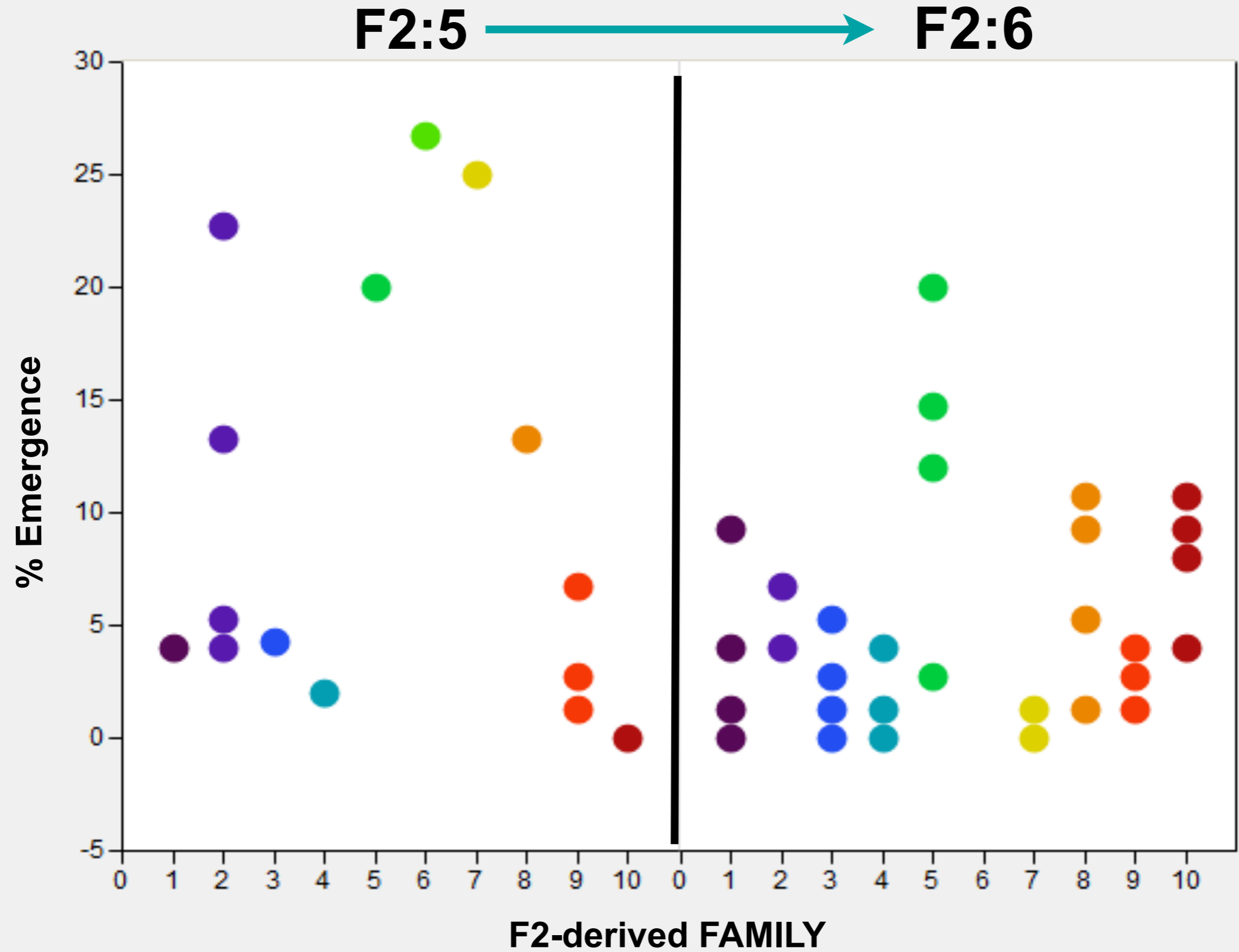
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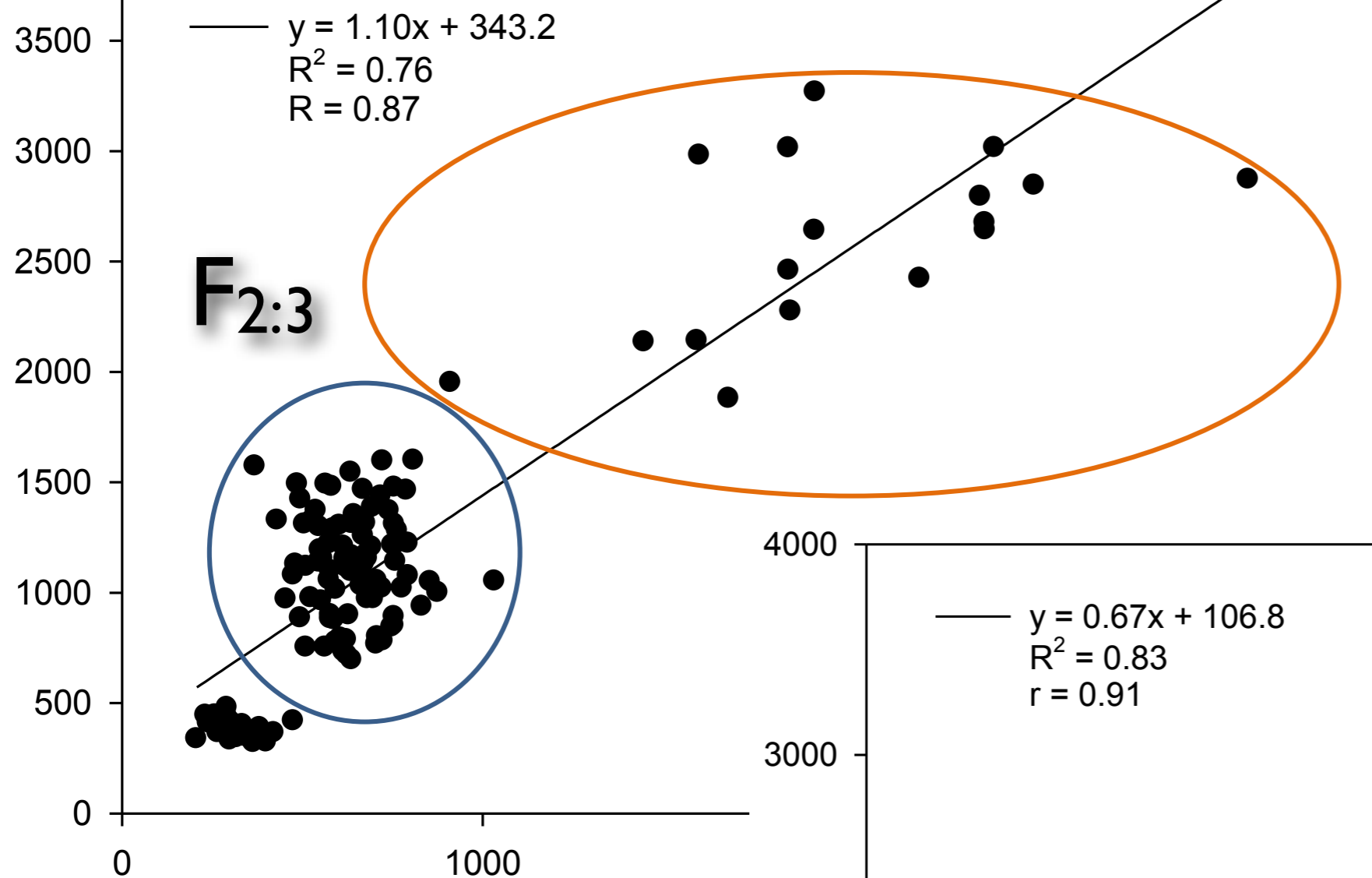
natural selection

LP populations = typical pops

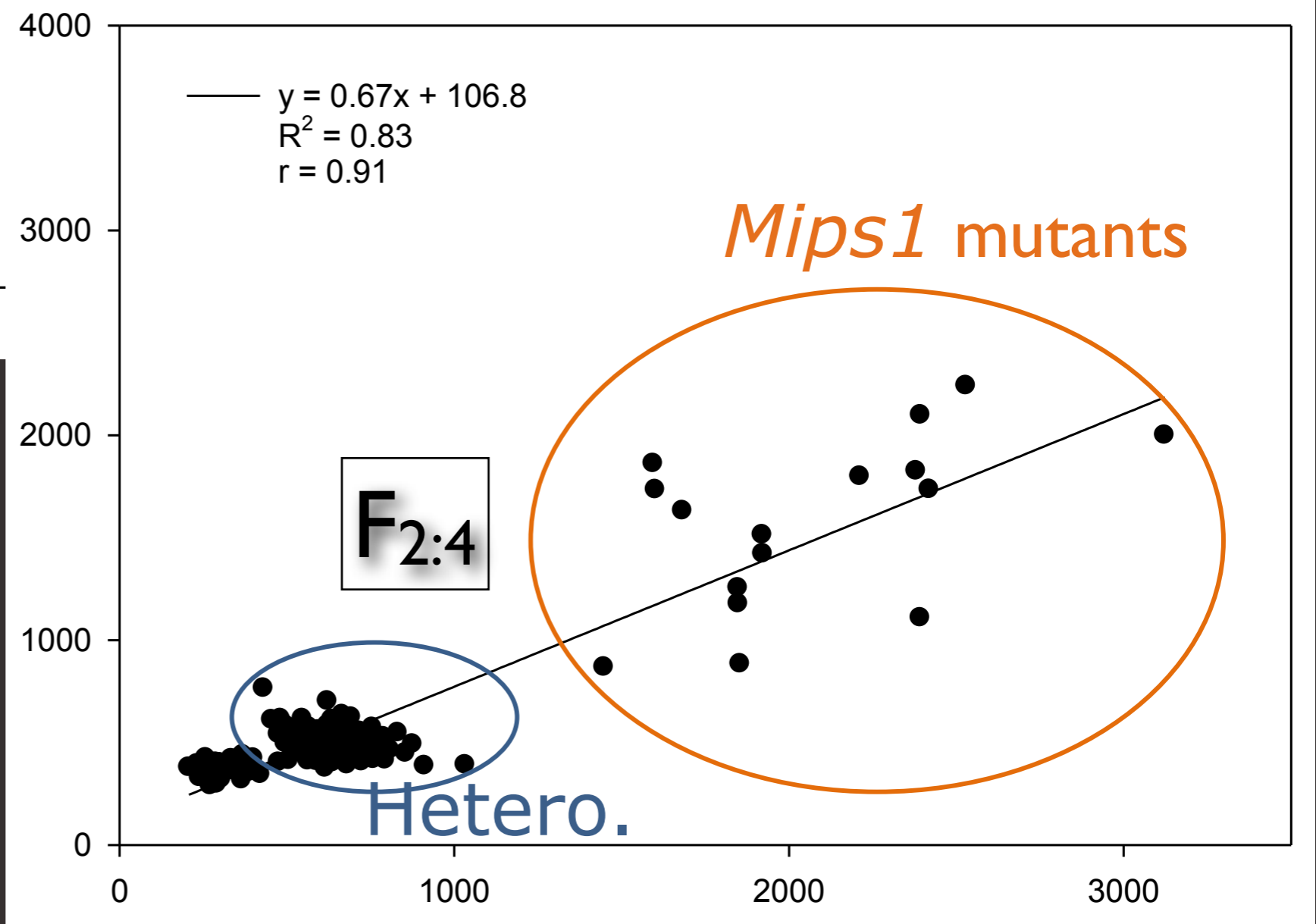
linkage

natural selection?



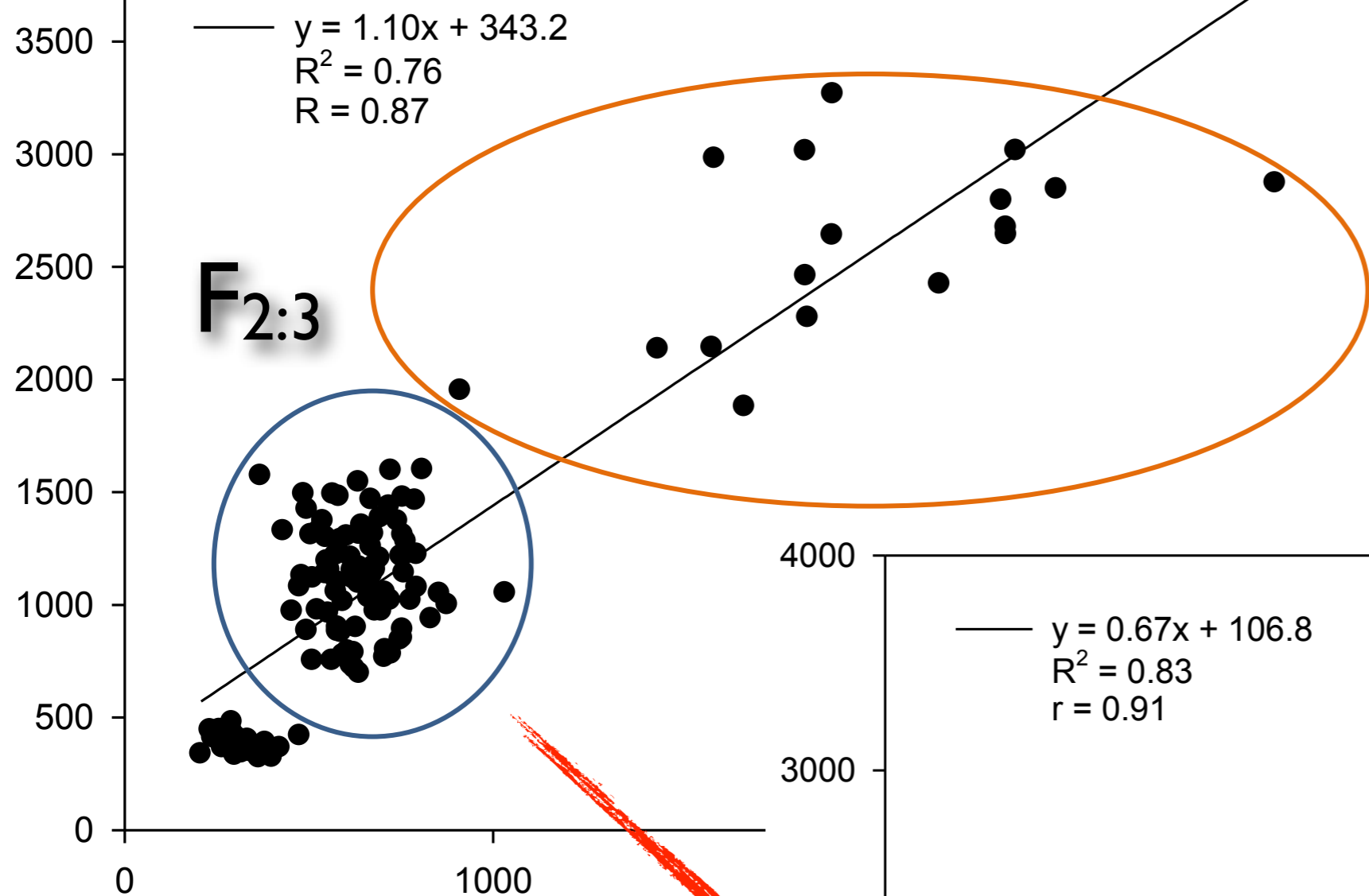


Shifting
Allele
Frequency

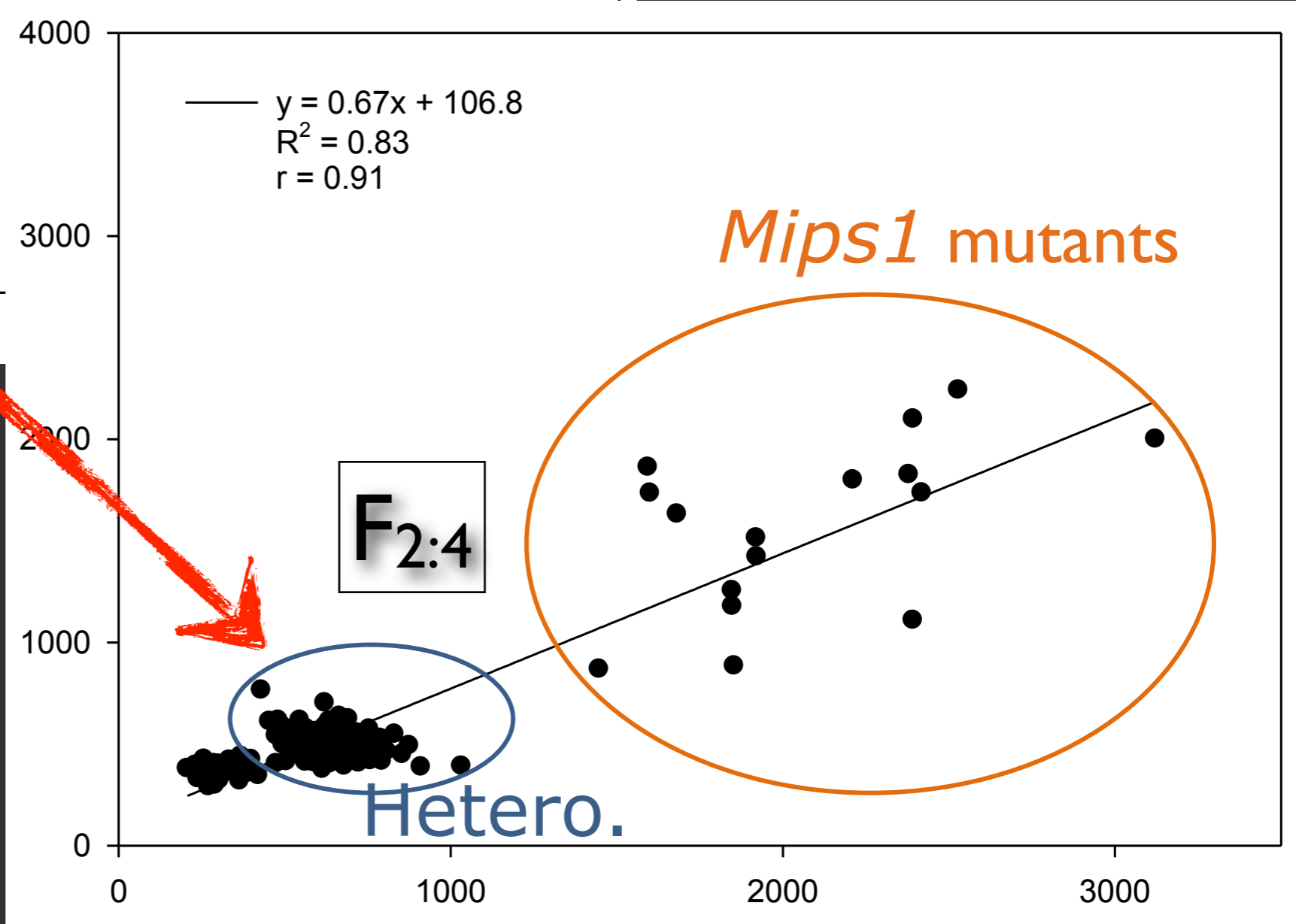


Maupin (2010)

F₂ Inorganic Phosphorus ($\mu\text{g g}^{-1}$)



Shifting
Allele
Frequency



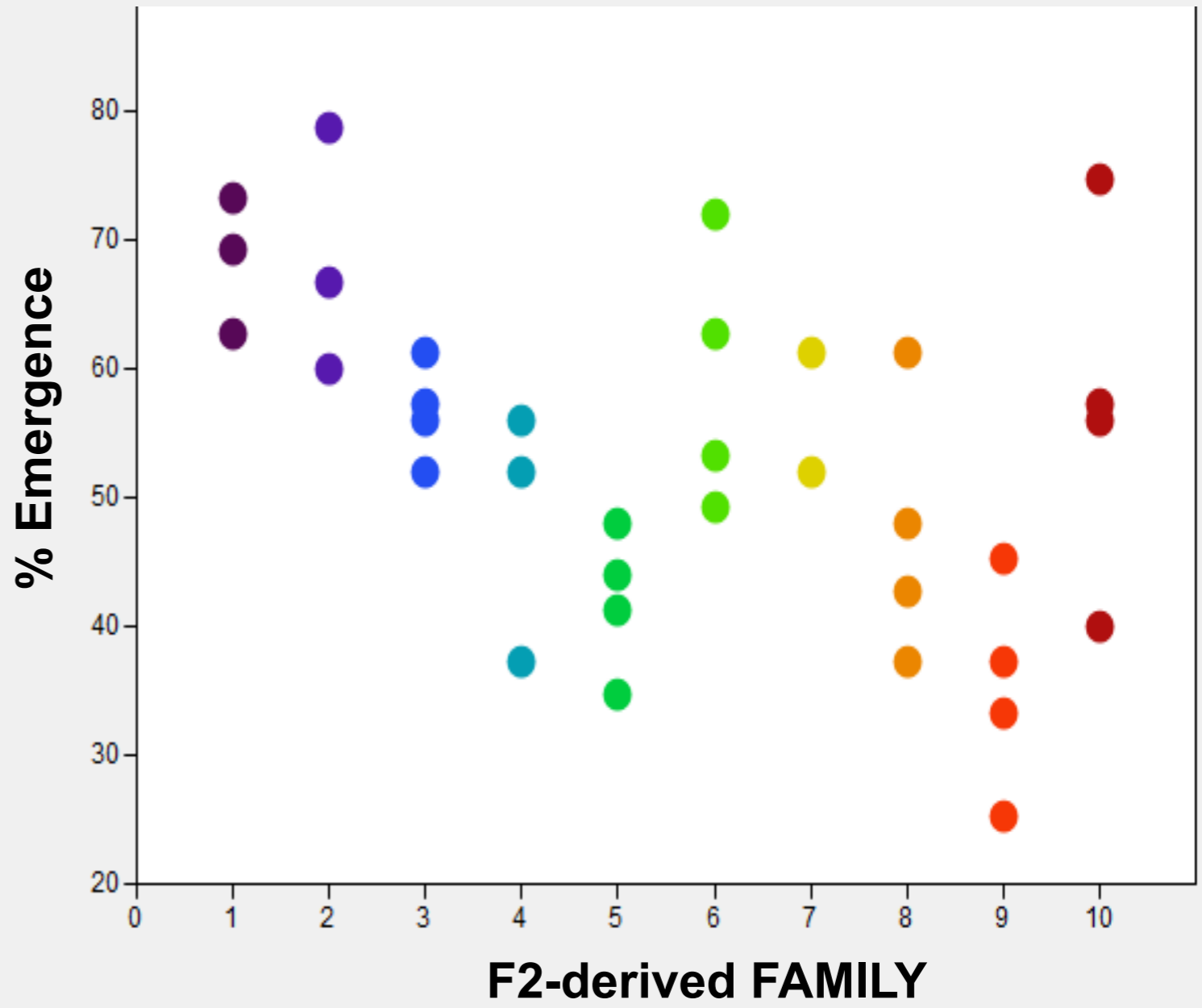
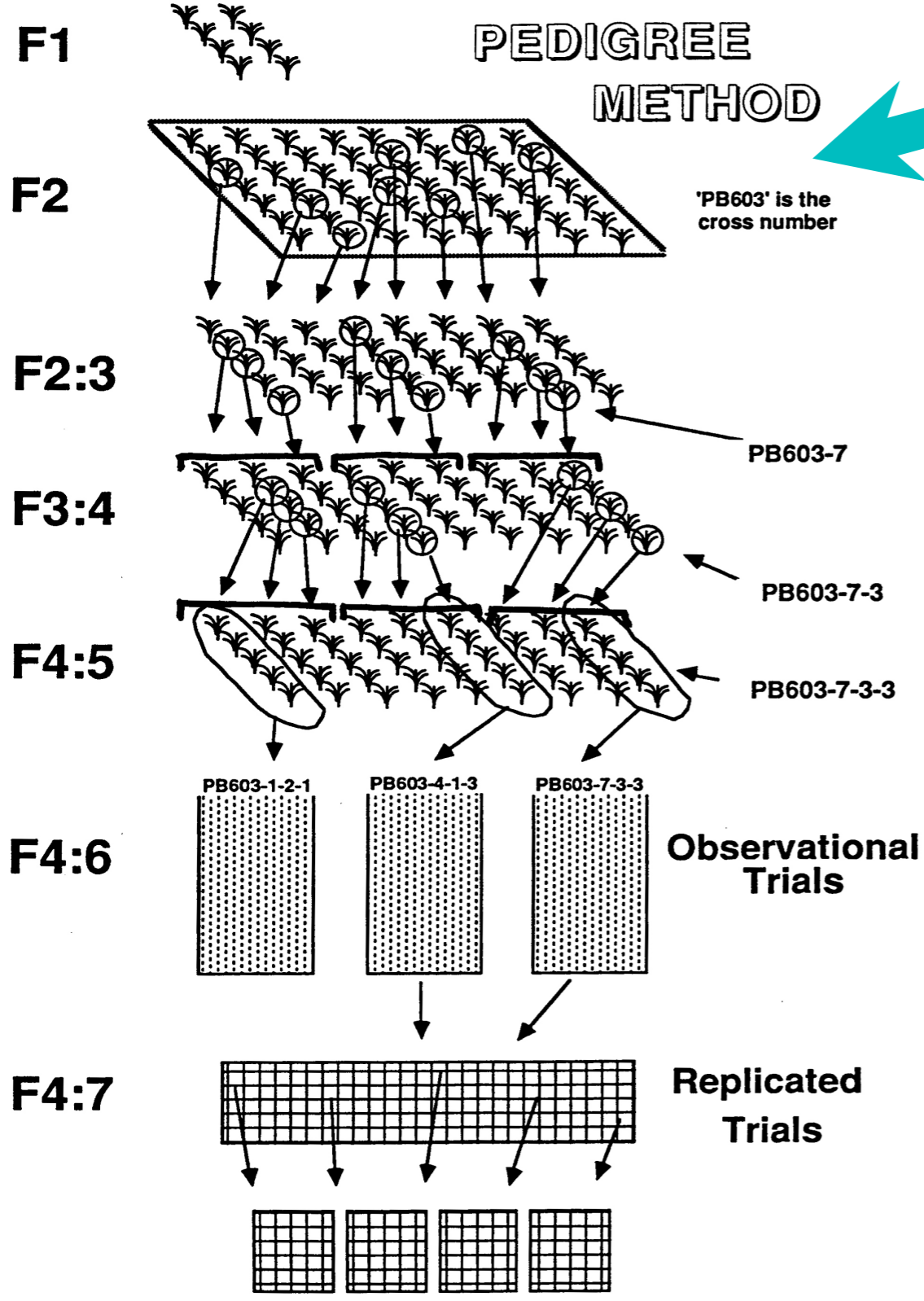
Maupin (2010)

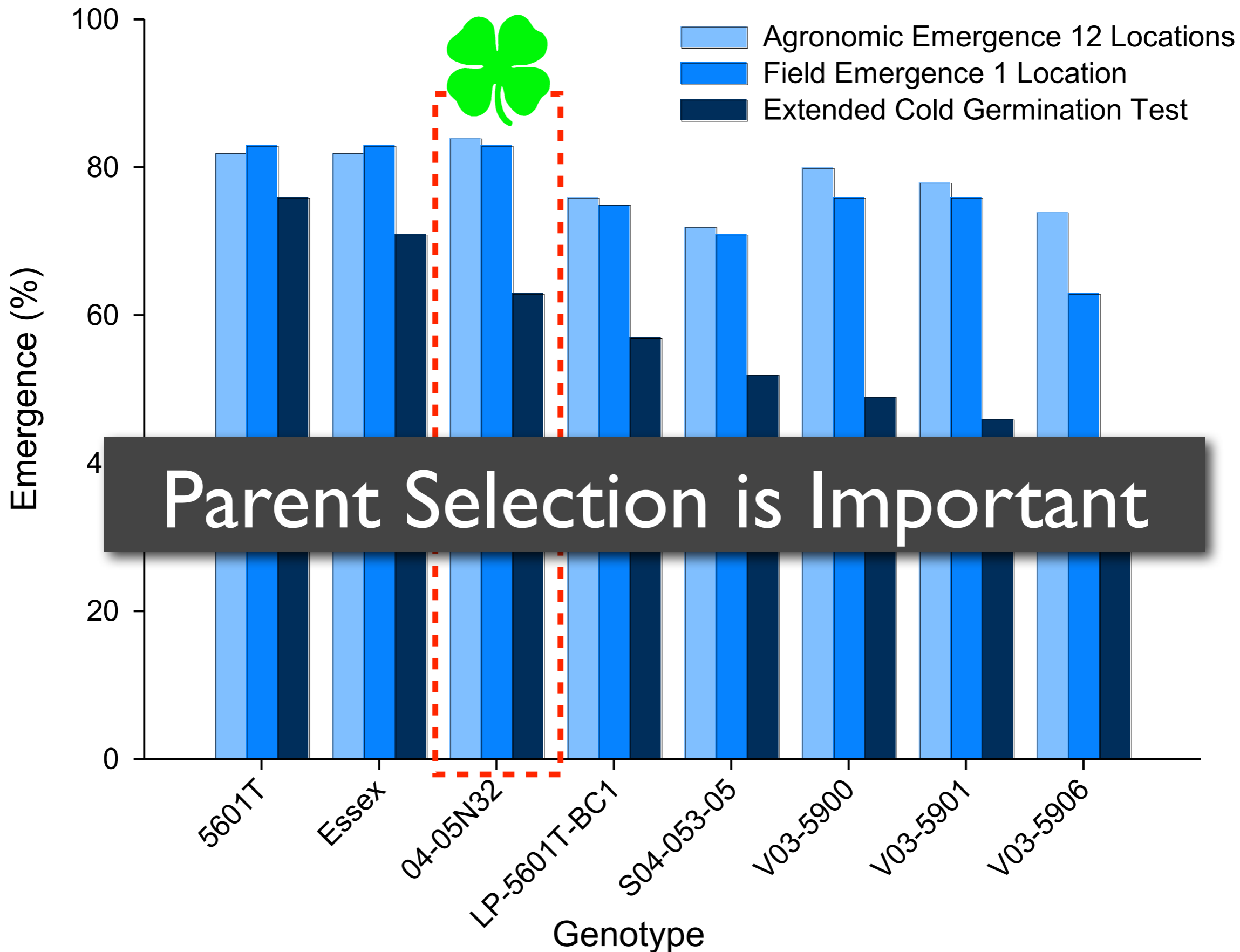
F₂ Inorganic Phosphorus ($\mu\text{g g}^{-1}$)

Pedigree Method: Genotypic Selection

“if it quacks 3 times its a duck”

Need large F₂

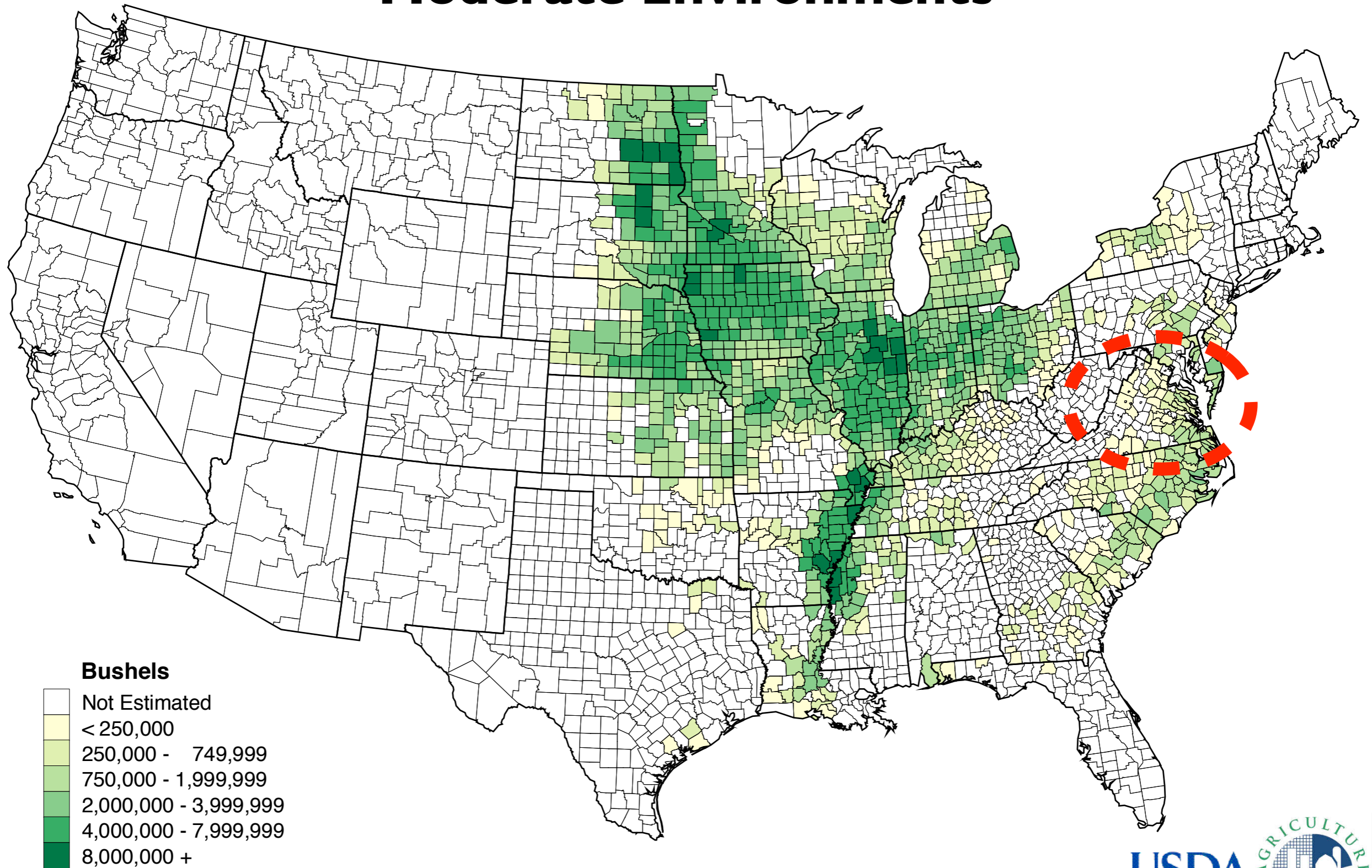




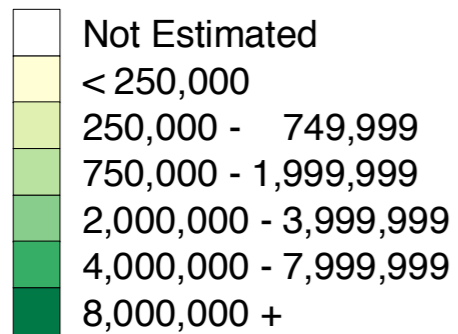
Parent Selection is Important

NAME	Phenotype	MG	2010 GERM TESTING		
			FIELD EMERGENCE	GERM ASSAY	ACCELERATED AGING
V07-6412	LP/ <i>MIPS1</i> mutant	M5	80	32	7
VS07-1779	LP/ <i>MIPS1</i> mutant	L4-E5	77	89	24
VS07-1265	LP/ <i>MIPS1</i> mutant	2-E3	27	67	30
VS07-1314	LP/ <i>MIPS1</i> mutant	M5	85	.	42
SSRT 3860	normal P	3	73	66	62
PI 588026A	normal P	4	71	83	66
0405N32	LP/ <i>Lpa</i> mutant	6	76	57	67
V03-4660	normal P	L4-E5	.	85	79
PI 588030	normal P	4	68	89	86
Glenn	normal P	M5	82	96	90
5002T	normal P	E5	64	.	.
MEAN			70	74	55

Moderate Environments



Bushels



Low Phytate Soybeans: *Can we solve the germ problem?*

- ▶ Low phytate soybeans can have poor G&E.
- ▶ Not all LP lines have poor G&E.
- ▶ Natural selection may not improve G&E in early generations.
- ▶ Pedigree selection may improve G&E in LP soy pops.
- ▶ Parents should contribute high G&E potential.
- ▶ Mid-Atlantic may be good environment for production of LP soybeans.

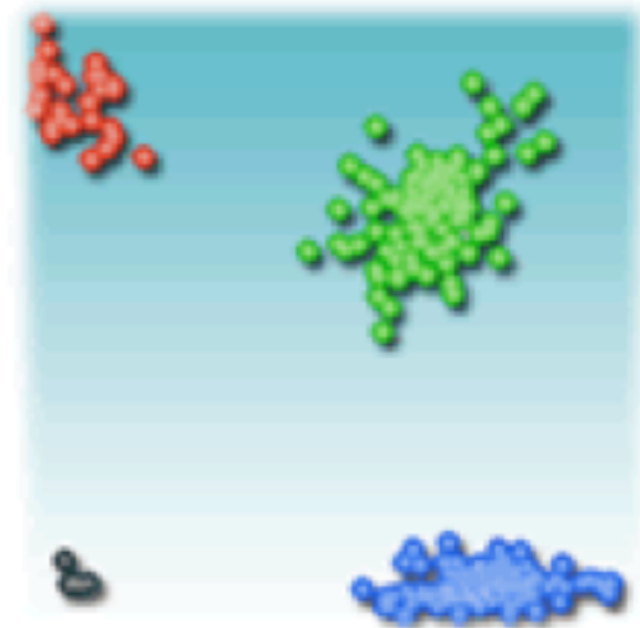
New Colorimetric Phytate Method

	Phytate		Phytate	
	---- $\mu\text{g P g}^{-1}$ ----		---- $\mu\text{g P g}^{-1}$ ----	
Genotype	Fe Method Mean	Fe Method Rank	HPIC Method Mean	HPIC Method Rank
S04-053-05	1032	1	860	1
04-05N32	1182	2	1137	2
5601T-BC1	1406	3	1282	3
V03-5906	2272	4	1887	4
V03-5901	2344	5	2092	5
V03-5900	2387	6	2058	6
5601T	3284	7	2897	7
Essex	3489	8	2947	8

KASPar SNP Assays: *MIPSI, Lpa1, Lpa2*

KASPar SNP Genotyping System and KASPar Direct real-time PCR system

The KBiosciences PCR SNP genotyping system is a novel homogeneous fluorescent genotyping system. We utilise a unique form of allele specific PCR that is distinct and different to conventional ARMS (Patent pending). It has been developed at KBiosciences and is currently in use on a daily basis. It offers the simplest and most cost effective way to determine SNP genotypes in the laboratory.



Maupin, L.M., M.L. Rosso, C. Shang, and Katy M. Rainey. 2011. Genotype \times environment interaction and stability of phosphorus concentration across twelve environments in two soybean germplasm sources with modified phosphorus composition. *Crop Sci.* Accepted.

Maupin, L.M., and Katy M. Rainey. 2011. Seedling emergence of two soybean germplasm sources with modified phosphorus composition. *Crop Sci.* Accepted.

Maupin, L.M., M.L. Rosso, and Katy M. Rainey. 2011. Environmental effects of soybean with modified phosphorus and sugar composition. *Crop Sci* 51:1-9.

Burleson, S., C. Shang, L.M. Maupin, L.M. Rosso, and K.M. Rainey. 2011. A High-Throughput Method of Phytate Detection. *Crop Sci.* *In Prep.*

Rosso, M.L., S. Burleson, L. Maupin and Katy M. Rainey. 2011. Development of Breeder Friendly Markers for Selection of Low Phytate Soybeans. *Molecular Breeding.* *Submitted.*

Acknowledgements

▶ Funding

- ▶ United Soybean Board
- ▶ Virginia Crop Improvement Research Foundation

▶ Germplasm and Locations

- ▶ V. Pantalone and lab
- ▶ G. Shannon and lab
- ▶ J. Burton and lab
- ▶ Schillinger Genetics
- ▶ Eastern VA AREC

▶ Seed Composition Assays

- ▶ Pantalone lab, Univ. Tenn.
- ▶ Dr. Chao Shang, VA Tech
- ▶ Dr. Rory Maguire, VA Tech
- ▶ ARS, Raleigh

▶ Effort

- ▶ Dr. Laura Maupin
- ▶ Dr. Luciana Rosso
- ▶ Dr. Saghai Maroof
- ▶ Sarah Burleson
- ▶ Tom Pridgen
- ▶ Lin Barrack
- ▶ Ted Lewis
- ▶ Melissa Woolard
- ▶ Ben Fallen
- ▶ Billy Rhodes

